x-ray imaging at the nanoscale / mit Röntgenaugen im Nanokosmos unterwegs



Tim Salditt, Institut für Röntgenphysik joint work with Matthias Bartels, Robin Wilke, Marius Priebe Klaus Giewekemeyer, Markus Osterhoff Universität Göttingen, Physik-Kolloquium Oldenburg, 7.11.2011



Content

- 1. Why x-ray imaging ?
- 2. The phase problem and its solution by iterative phasing
- 3. far-field (CDI) versus near-field (propagation) imaging
- 4. The Göttingen endstation for nano-imaging at the coherence beamline PETRAIII HASYLAB /DESY
- 5. X-ray waveguide optics
- 6. Applications in biological imaging
- 7. The advent of X-ray Free Electron Lasers (XFEL)

joined work with Matthias Bartels, Robin Wilke, Marius Priebe, Markus Osterhoff, Sebastian Kalbfleisch Scen Krüger, Henrike Neubauer, Dr. Klaus Giewekemeyer



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absorption versus phase contrast

 $U(X,Y) = U_0(X,Y) \quad \tau(X,Y)$

$$n=1-\delta+i\beta$$

x-ray index of refraction

$$\tau(x, y) = \exp[-k \int_{z}^{z+d} dz \left(\beta(x, y, z) + i \delta(x, y, z)\right)] \qquad \qquad \delta = 10^{-5} .. 10^{-9} \\ \beta = 10^{-7} .. 10^{-13}$$



 $\delta >> b$ for low Z elements and high photon energies \rightarrow phase contrast

Propagation imaging



P. Cloetens et al. et al. ,1999; S.Wilkins et al. , 2000-2004

Density in 3D : resolution AND contrast matter !



Cochlea: Phase contrast vs. Absorption contrast

X-ray advantage No.2: (a) quantitative contrast and weak scattering cross section



index $n = 1 - \delta + i \beta$, δ , $\beta \approx 10^{-5-}10^{-6}$ close to 1.

⇒ reduced reflections at internal interfaces *no multiple scattering*

(look through foam of beer, R.W. Pohl 1939)



(Born Approx.)

crystallography = averaging over 10¹² - 10¹⁸ copies of the same molecule









 $\sum e^{i\vec{q}\cdot\left(n_{1}\vec{a}+n_{2}\vec{b}+n_{3}\vec{c}\right)}\Big|^{2}$ 12 $\sum_{n=1}^{N} e^{i\vec{q}\cdot\vec{r_n}} \Big|^2$ $S(\vec{q}) =$ unit cell $n_1 n_2 n_2$



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Advantage No.3: small wavelength – high resolution



The phase problem Amplitude

Phase



 $\widehat{\rho}_{kl} = \frac{1}{\sqrt{N \cdot M}} \sum_{i=0}^{N-1} \sum_{j=0}^{M-1} \rho_{ij} e^{-2\pi i \left(\frac{ik}{N} + \frac{jj}{M}\right)}$



Fresnel Zone Plate (FZP)

"Oversampling": a strategy to render the problem unique ?



Information theory (Shannon) ? really more information

"Oversampling"

$$I(q_{kl}) = F(q_{kl}) F^{*}(q_{kl}) = \left| \sum_{i}^{N-1} \sum_{j}^{N-1} \rho_{ij} e^{-i(q_{k} x_{i} + q_{l} y_{i})} \right|^{2}$$

h, k, i, j = 0...N - 1
$$F(q) = F^{*}(-q)$$

J.Miao et al., J. Opt. Soc. Am. '98 J.Fienup, Appl. Opt. 21, 2758 (1982) J.Miao, J. Kirz & D. Sayre, Acta Cryst. D 56, 1312 (2000)

"Oversampling": a strategy to render the problem unique ?



 $F(q) = F^*(-q)$

 $N^2\,$ unknowns $r_{ij\prime}\,$ but only $N^2/2\,$ independent equations

 $\rho(r) = 0$ out of support !

If support is half the size of the field of view in both both dimensions:

 $N^2\,$ unknowns $\rho_{ij},$ 4 $N^2\,$ equations 2 N^2 independent equations

J.Miao et al., *J. Opt. Soc. Am.* '98 J.Fienup, *Appl. Opt.* 21, 2758 (1982) J.Miao, J. Kirz & D. Sayre, *Acta Cryst.* D 56, 1312 (2000)

how to solve the nonlinear set of equations: iterative algorithms



$$\rho_{i+1}(r) = \begin{cases} \rho_i'(r) \ r \in S \cap \rho_i'(r) > 0\\ \rho_i(r) - \beta \rho_i'(r) \ r \notin S \cup \rho_i' \le 0 \end{cases}$$

J. Miao et al., Nature **400**, 342 (1999)

Ptychography: using the overlap constraint

 $z \gg d$ d diffraction diffraction pattern 4 pattern 1 $P(\mathbf{r})$ detector plane object plane FFT probe $\mathcal{Y}(r)$ FFT⁻¹ O(r)R. Wilke et al. (unpublished) diffraction diffraction pattern 3 pattern 2 fit Rodenburg et al. PRL 98, 034801 (2007) -0.2 -0.4 2.5 -0.6 ≎ ptychographia reconstruction with x-rays: Ē Rodenburg et al. PRL (2007) Ē -0.8 simultaneous reconstruction of illumination field: -1 [mm] Thibault et al. Science (2008) Guizar-Sicairos, Fienup, Optics Express (2008) Ξ application to biological cells: -1-0.50 Φ [rad] -1 [mm] Klaus Giewekemeyer et al., PNAS (2010) (b)

 $\Psi(\mathbf{r}) = P(\mathbf{r} + (0, 0, d)) \cdot O(\mathbf{r}) \qquad |\mathcal{F}(\Psi)|^2 = |\mathcal{F}(P_d) \times \mathcal{F}(O)|^2$



scanning and diffraction



Maximum spacial frequency given by detector size and distance (Pilatus pixel det.) $15/\mu m$ (d = 67nm), possible, but noise issues (Poissonian noise)!

Giewekemeyer et al, PNAS 2010



DNA packing in nucleoids: *Deinococcus Radiodurans*



• Among most radiationresistant organisms on earth, can survive 15 kGy of ionizing radiation



Right: super-resolution by phasing the coherent diffraction pattern

- Freeze-dried, unstained and unsliced cells
- Overall phase shift of single cell 0.25-0.3 rad (< 10% p), consistent with simulations
- 2500 iterations of SXDM algorithm, averaged over each 5th iterate, starting at 2000

f [rad] 0 -0.05 -0.1 -0.15 -0.2 -0.25 -0.3 -0.35 -0.4



TEM-slices, Os-stained chromatin Levin-Zaidman, Science (2003)



2x3 m size, tetrad morphology 4 identical copies of the genome

Chemical Contrast in Soft X-Ray Ptychography

Mike Beckers, 1,* Tobias Senkbeil, 1 Thomas Gorniak, 1 Michael Reese, 2 Klaus Gieweker $^{---}$ Tim Salditt, 3 and Axel Rosenhahn 1,5,†

Soft x-rays (water window): Absorption and phase Carbon K-edge -> chemical contrast



G FIG. 2 (color). Reconstructions of amplitude and unwrapped phase (in radians) of five ptychographic data sets recorded at different energies around the measured C = O absorption peak of PMMA. The sample, a mixture of five PMMA beads and four SiO₂ beads each 2 μ m in size, shows absorption and phase shift



FIG. 3 (color). *D. radiodurans* cells. (a) Bright field microscopy image of a part of the specimen. Marked is the area of the ptychographic scan used for reconstruction. (b) Reconstructed amplitude of the cells. (c) Unwrapped phase shift of the cells and (d) detailed view.

Hard x-rays : Absorption neglegible large penetration and focal depth -> tomography



R. Wilke et al. (unpublished)

illumination function: plane wave, curved, structured,



Far-field and near-field diffraction patterns !



Fresnel scaling theorem: an equivalence between parallel and point source illumination



hologram recorded with the point source corresponds to a hologram recorded with a plane wave at an effective defocusing distance



→magnification allows for a spatial resolution below detector pixel size!
 → plane wave setup used for simulations and reconstruction

Imaging formation: Fresnel diffraction integrals



transformation from spherical to parallel beam, z-> z_{eff} M magnified coordinate system normalisation of the measured intensity I by empty beam

$$\overline{I}(x,y) \simeq |\psi_{\rm in} D_{z_{\rm eff}}[\chi(x,y)]|^2 / |\psi_{\rm in}|^2$$
$$= |D_{z_{\rm eff}}[\chi(x,y)]|^2.$$



 $I(x,y) = |D_{z_{\text{eff}}}[\psi_{\text{in}}\chi(x,y)]|^2 \simeq |\psi_{\text{in}}D_{z_{\text{eff}}}[\chi(x,y)]|^2$

Waveguide based propagation imaging schematic setup



- 1. Homogeneuos signal level in detection plane
- 2. High dose efficiency
- 3. Less scanning overhead for tomography (full field)

Waveguide based propagation imaging setup – P10 at PETRA III



Kalbfleisch et al. , AIP Conf. Proc., 2011

KB mirrors

mirror –precision polishing and metrology



Fig. 3. Hard X-ray focussing mirror for the holography end-station of beamline P10 at PETRA-III at the alignment set with three steel balls. The 10 mm thick substrate was finished on a $10 \text{ mm} \times 5 \text{ mm}$ long aperture section by EEM-finishing technology.

Table 1

P

S

E

Shape parameter of an elliptical cylinder mirror, for hard X-ray focussing at PETRA-III, as measured using BESSY-NOM and a Micromap Promap interference microscope (using magnifications $2.5\times,20\times$ and $50\times$).

Specification	Measurement results
85 500	85 500
200	200
4.0	4.0499
Not defined	42 648.2
	0.13 µrad rms
3 nm pv	1.0 nm rms/3.1 nm pv
≤ 0.15 nm rms	$\begin{array}{l} S_{\rm q} = 0.135 - 0.164 \mbox{ nm rms} (2.5 \times) \\ = 0.107 - 0.122 \mbox{ nm rms} (20 \times) \\ = 0.101 - 0.122 \mbox{ nm rms} (50 \times) \\ S_{\rm a} = 0.106 - 0.126 \mbox{ nm rms} (2.5 \times) \\ = 0.082 - 0.096 \mbox{ nm rms} (20 \times) \\ = 0.079 - 0.093 \mbox{ nm rms} (50 \times) \end{array}$
	Specification 85 500 200 4.0 Not defined 3nm pv ≤ 0.15 nm rms



Fig. 1. Principal set-up of a mirror based moving penta-prism slope measuring profiler as realized for BESSY-NOM.





Fig. 4. HardX-ray focussing mirror for the holography end-station of beamline P10 at PETRA-III. Left—profile of residual slope measured at two different alignment conditions from side A to B and after a 180° rotation from side B to A. For better visualization a shift of 0.2 µrad was added to the profiles. Right—the corresponding profiles of residual height, gained by integration of the slope data.

Elastic emission machining and Interferometry (Yamauchi group, Osaka University Commercialized by JTEC Corp, Osaka Metrology: F. Siewert et al., NIM A 2011

measured intensity distribution compared to simulation



T. Salditt et al. Opt. Express 2011









measured coherence function in good agreement with simulation

T. Salditt et al. Opt. Express 2011







Osterhoff, Salditt NJP 2011

New generations of x-ray waveguides: (a) crossed high transmission planar waveguides (b) lithographic channels in Si sealed by wafer bonding



T. Salditt, S. Krüger, C. Baehtz, Phys. Rev. Lett. 2008; S. Krüger et al., Opt. Express 2010; H. Neubauer, M. Kanbach et al., unpublished

Waveguide based propagation imaging

Waveguide system



Calculation of near-field intensity distribution from farfield diffraction pattern: *sub 15 nm beam size in two dimensions*



beam size (FWH		
reconstruction:	9.2nm x 9.6 nm	
simulation:	12.5nm x 13.6 nm	
autocorrelation:	14.2nm x 17.9 nm	

S. Krüger et al., Optics Express, 2010

world record in one-dimension: 7nm focus by EEM polished mirror @1km long beamline 29 Spring8 (Yamauchi group, Osaka Univ.)

H. Mimura *et al.*, Nature Physics, 2010 *Breaking the 10 nm barrier in hard x-ray focusing*

X-ray imaging of cells: *Dictyostelium Discoideum* -0.04 5 -0.08 10 hm -0.12 1.08 15 1.04 -0.16 20 -0.2 0.96 5 10 15 20 0.92 1.08 ^{µm} z1=3mm Waveguide flux = 1e8 ph/s M=1763 t = 60 x 0.5 sec = 30 sec p=32 nm 1.04 t=150 s 1 0.96 M. Bartels, M. Priebe et al., unpublished



ID 22NI, 17.5 keV 35 x 35 nm crossed WG 8.3 mm defocus





Zellen (*Dictyostelium Discoideum, cryo. fixiert & gefrier* getrocknet)

- Phaseninformation durch Referenzwelle
- quantitativer Dichtekontrast
- •iterative Rekonstruktion
- •Lösung des Zwillingsbildes (hologr. Artifakt)

K. Giewekemeyer ,S. Krüger, S. Kalbfleisch, M. Bartels, C. Beta, T. Salditt , Physical Review A, 2011



Propagator
$$D_z = FFT^{-1} \exp[iz \sqrt{k^2 - k_x^2 - k_y^2}] FFT$$

$$P_{D} :' \text{ projector'} \quad \text{in detector} \quad \text{plane}$$

$$\left| \widetilde{\chi}_{n}(x, y) \right|^{2} = \left(1 - \frac{D}{d} \right) \overline{I}(x, y)$$

$$+ \frac{D}{d} \left| \widetilde{\chi}_{n}(x, y) \right|^{2}$$

$$d^{2} = \frac{1}{N} \sum_{x, y} \left(\left| \widetilde{\chi}_{n}(x, y) \right|^{2} - \overline{I}(x, y)^{2} \right)^{2}$$

$$D = \sqrt{2 I} < I_{0} > \text{noise}$$

 $P_{s}: \text{projector} \quad \text{in sample plane} \\ \left| \chi_{n+1}(x,y) \right| = \left| \chi_{n}(x,y) \right| - \beta \left(\left| \chi_{n}(x,y) - 1 \right| \right) \\ \arg(\chi_{n+1}(x,y)) = \left\{ \begin{array}{c} \arg(\chi_{n}) - \gamma \arg(\chi_{n}) & \forall (x,y) \notin S \\ \min(\arg(\chi_{n},0) & \forall (x,y) \in S \end{array} \right. \end{cases}$

S support (from holographi c reconstruc tion)

X-ray imaging of bacterial cells Deinococcus Radiodurans





M.Bartels, M. Priebe et al., submitted.

DNA packing in nucleoids: Tomography of *Deinococcus Radiodurans*

- Among most radiationresistant organisms on earth, can survive 15 kGy of ionizing radiation
- Very effective DNA repair mechanism, DNA packing debated



2x3 μm size, tetrad morphology 4 identical copies of the genome

Matthias Bartels et al. (unpublished manuscript)



Three-Dimensional Visualization of a Human Chromosome Using Coherent X-Ray Diffraction



FIG. 2 (color). Coherent diffraction pattern of an unstained human chromosome and its reconstructed projection image. The



K. Giewekemeyer et al., PNAS 2010

The advent of x-ray free electron lasers 9 orders in peak brilliance !



Figure 1. Peak brilliance of storage ring and FEL sources.

editorial

X-ray vision

The century-old field of X-ray physics is being rejuvenated by new forms of ultrabright sources based on laser technology, promising a revolution in imaging capabilities.

This month *Nature Photonics* presents a special Focus Issue dedicated to the latest advances in X-ray optics, a field that has a long and rich history.

"I have seen my death," exclaimed Anna Röntgen in mid-November, 1895. She had just seen the first ever human X-ray image — a picture of her own hand revealing the bones beneath the flesh.



setting, using either high-harmonic generation, laser wakefield or various types of plasma sources. This is in stark contrast with the immobile nature of synchrotrons, i in which most high-resolution work has been done so far. X-ray imaging schemes generally utilize soft X-rays, with or without lenses, in a modality that is



Simulated diffraction images of a single molecule in different representations (a)+(b) and at different intensities (c)-(f).



C10 (Salditt /Köster) Single pulse cellular imaging (high throughput)







 $N_0 = 10^{12}$ photons / pulse assume $N_c = 10^5$ per resolution element r $\rightarrow 3000x3000$ pixel in field of view V $\lambda = 0.1$ nm, V = 10µm, $z_1 = 10$ mm, r = 3nm focus $\Delta = 100$ nm fluence at defocus $N_0 \lambda^2 \Delta^2 / z_1^2$ 3D information from anlylography ?

 $\rightarrow @ 10^8 W/cm^2$: 100mV transmembrane Voltage



FLASH experiments : D.D. Mai, T. Reusch, J. Hallmann, TS Vartaniants, A. Mancuso, A. Rosenhahn, A. Singer, J. Gulden, O. Yefanov T. Senkbeil, T. Gorniak, M. Beckers, Rolf Treusch, Stefan Düsterer

the team / collaborations: acknowledgements

waveguide holography, tomography, neural cells and tissues
phase reconstruction algorithms, ptychography
phase reconstruction algorithms, ptychography, cellular imaging
cellular imaging and diffraction
z multicellular organisms
waveguide microscope
ven Krüger waveguide optics and fabrication
numerical optics, focusing, mirror design
mann, Tobias Reusch FLASH experiments

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A. Rosenhahn, U. Heidelberg
M. Sprung, I. Vartanyants, S. Düsterer. R. Treusch DESY
H. Metzger, R. Tucoulou, P. Cloetens ESRF
F. Pfeiffer, A. Diaz, C. Kewish, P. Thibault SLS/ TU München

